

1 SHORT TITLE: LEARNING AND OVIPOSITION

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3 The role of learning in the oviposition behavior of the silkworm moth (*Bombyx mori*)

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Abstract

One of the most relevant behaviors in the life of some insects is oviposition, because reproductive fitness largely depends on the choice of egg-laying site. We report one experiment that explored the influence of Pavlovian conditioning on oviposition choices of the silkworm. Our results show that moths that jointly experienced an odor (conditioned stimulus) and mulberry leaves (the preferred oviposition place for the moths) preferred to lay their eggs near the odor when it was present, whereas moths in which the odor and the mulberry leaves never appeared together showed no preference. This result provides evidence of the important role that a psychological process such as Pavlovian conditioning has for the survival of this species.

Key words: silkworm; lepidopteran; Pavlovian conditioning, oviposition behavior.

47           Although there are 80 years of study of learning in insects (e.g., Thorpe & Jones,  
48 1937), and such research is growing, research in insect learning is still a minority.  
49 Animals must adjust to changing conditions, and they benefit from learning in variable  
50 environments with some predictability by using information they have previously  
51 acquired. The ability to change behavior after experience is adaptive (e.g., Alcock  
52 1993). The experiment presented here provides new evidence about the learning  
53 capabilities of silkworms and the functional advantages obtained by learning in this  
54 species.

55           Scientific evidence has shown the learning capabilities of insects for some crucial  
56 behaviors such as searching for food (e.g., Cunningham, Moore, Zalucki & West, 2004;  
57 Dukas & Bernays, 2000; Riffell et al, 2008), mate finding (e.g., Anderson et al., 2007;  
58 Anderson, Sadek, Larsson, Hansson & Thöming, 2013; see Dukas 2006 for a review)  
59 and rearing offspring (e.g., Cunningham, Jallow, Wright & Zalucki, 1998; Olsson,  
60 Anderbrant & Lofstedt, 2006). For instance, Cunningham et al. (2004) explored whether  
61 learning influences innate preferences for two odors, phenylacetaldehyde and  $\alpha$ -pinene,  
62 in the nectar-feeding moth *Helicoverpa armigera*. In a preference test, prior to a  
63 Pavlovian conditioning procedure, moths flew towards both odors, with a preference for  
64 phenylacetaldehyde. During conditioning, a group of moths was exposed to one of the  
65 two odors while feeding on a sucrose solution. Two control groups were used for  
66 comparison. One group of moths was exposed to odors without allowing feeding while  
67 another received no exposure to either the odors or feeding. During the test, moths  
68 could choose between the odors. Moths that received the odors paired with feeding  
69 showed a clear preference for the odor associated with the sucrose, whereas this change  
70 in preference was not seen in the controls. In the moths' natural environment this

71 learning would allow them to discriminate odors that signal more nutritious feeding  
72 sources.

73 This kind of result highlights the relevance of learning for the species' survival. In  
74 that theme, other studies have evaluated the adaptive advantage of an associative  
75 learning process, such as Pavlovian conditioning, in a variety of taxa such as birds  
76 (e.g., Adkins-Regan & MacKillop, 2003; Domjan, Blesbois, & Williams, 1998), fish  
77 (Hollis, Dumas, Singh & Fackelman, 1995; Hollis, Pharr, Dumas, Britton & Field,  
78 1997), mammals, (Coria-Avila, 2012; Zamble, Hadad, Mitchell & Cutmore, 1985) and  
79 insects (Dukas, 1998; Dukas & Bernays, 2000; Guillette, Hollis & Markarian, 2009).

80 In an experiment with grasshoppers, Dukas & Bernays (2000) showed the  
81 importance of associative learning for optimal growth. These authors arranged two  
82 groups of grasshoppers so that the experimental group was exposed to a combination of  
83 cues (spatial location, taste, and color) paired with a balanced diet including a suitable  
84 proportion of carbohydrates and protein that allows maximal growth, or a deficient diet  
85 with no carbohydrates. A control group was unable able to use any of the cues for  
86 choosing between the diets. The results showed that grasshoppers in the experimental  
87 group were able to use the predictive cues to visit the balanced diet more frequently and  
88 ate more of the balanced diet. Hence, the growth rate was higher in experimental than in  
89 control group.

90 Oviposition is one of the most important behaviors for a lepidopteran since  
91 choosing the substrate upon which to lay eggs could be momentous for the survival of  
92 the offspring. New larvae will be properly fed when they are born surrounded by food  
93 without the need to move for foraging. The success of oviposition in the silkworm not  
94 only depends on the substrate. There are a number of neural, chemical, physical,  
95 environmental, and behavioural factors involved in this behavior. For instance, the

96 number of eggs laid and the time the moth spends ovipositing are influenced by  
97 environmental conditions such as temperature, light, humidity, and surface texture.  
98 Once the mating is complete and the female moths have found a suitable place, 90% of  
99 the eggs are deposited within a 24-hour period after mating. Several researchers have  
100 established that higher oviposition rates occur at a temperature of about  $25 \pm 1^\circ \text{C}$  and 80  
101  $\pm 5\%$  relative humidity (e.g., Singh & Saratchandra, 2004).

102       There are multiple examples about how larval or oviposition experiences  
103 influence later oviposition choices. However, the evidence regarding associative  
104 learning in adult lepidopterans in relation to oviposition is scarcer. For instance, several  
105 studies state that female moths learn associations between leaf shapes and the chemical  
106 compositions required for oviposition (Papaj, 1986), or between the appearance of  
107 oviposition substrates and the chemical oviposition stimulants (Traynier, 1984, 1986).  
108 Lepidopterans and other insects show an innate preference for a given chemical  
109 composition of the substrate. Thus, the plant odor is a valuable cue for this kind of  
110 animal to choose their oviposition substrates. For example, Traynier (1986) tested the  
111 preference of cabbage butterflies for laying their eggs on discs with different sizes and  
112 colors that had been wetted with either a sinigrin solution or water. Sinigrin is a  
113 chemical compound in plants like cabbage that releases the oviposition behaviour in  
114 lepidopteran species such as the cabbage butterfly. The results showed the individuals  
115 preferred to lay the eggs on discs with a similar appearance to those containing the  
116 sinigrin solution.

117       As far as we know, this study and the rest of studies about learning in oviposition  
118 behavior explore the insects' ability to associate some features of the plants with its  
119 suitability for oviposition. However, the experiment we present in this paper evaluates  
120 the silkworm's ability to learn to predict the presence of food through cues different

121 from the food itself. The silkworm only feeds during the larval period. In its adult  
122 phase, i.e., when it becomes a moth, it does not feed and survives on the energy reserves  
123 created during the larval state. Unlike other lepidopteran species, the silkworm is  
124 monophagous, it solely eats mulberry leaves. Therefore, *a priori* the silkworm does not  
125 need to learn to choose the most suitable food. Nevertheless, there would still be an  
126 adaptive advantage to be able to learn that specific stimuli can predict the presence of  
127 mulberry leaves. Such a capacity would allow moths to better locate food sources upon  
128 which to lay eggs.

129         A moth's capacity for Pavlovian conditioning has been clearly demonstrated  
130 (see as well Daly & Smith, 2000; Fan, Anderson & Hansson, 1997; Jørgensen,  
131 Strandén, Sandoz, Menzel & Mustaparta, 2007). Based on those findings, it can be  
132 expected that the preference for oviposition can also be conditioned. Thus, our goal  
133 in this experiment was not to explore the influence of previous experience for  
134 oviposition, but to specifically test if moths would prefer to oviposit near a stimulus  
135 that had been previously associated with the host plant.

136         The general frame for the experiment presented here is the Hopkins' host-  
137 selection principle (HHSP). The principle states that "a species which breeds in two  
138 or more hosts will prefer to continue to breed in the host to which it has become  
139 adapted" (Hopkins, 1916, p. 353). Related to the HHSP, the natal habitat preference  
140 induction hypothesis (NHPI) more specifically states that insect females prefer to  
141 lay their eggs on the host species on which they developed as larvae (Davis &  
142 Stamps, 2004). Several studies have supported these ideas (e.g., Blackiston, Casey,  
143 & Weiss, 2008; Gandolfi, Mattiacci, & Dorn, 2003; Tully, Cambiazo, & Kruse,  
144 1994; but see e.g. Barron, 2001; Janz, Söderlind & Nylin, 2008 for results to the  
145 contrary). Hence, it could be expected that the silkworm moths will prefer to

146 oviposit on mulberry leaves, the plant they ate during their larval period. Moreover,  
147 if the oviposition of silkworms can be conditioned, when an odor (conditioned  
148 stimulus, CS) is paired with mulberry leaves (unconditioned stimulus, US), the  
149 moths will lay the eggs near the odor, similarly to way they do on mulberry leaves.

150 To test those hypotheses, a group of female moths in the “Paired Group”  
151 experienced an odor (CS) paired with a mulberry leaf unconditioned stimuli (US). Two  
152 control groups were used. One group, “Unpaired Group” experienced the CS and the  
153 US unpaired, so that the stimuli were never were presented together and the final “CS-  
154 Only Group” experienced only the CS. During the test, one half of each group was  
155 exposed to the US and the other half to the CS in a box with a male moth. We counted  
156 the number of eggs laid on the area in which the stimulus was presented (the Stimulus  
157 Zone) or on the rest of the box (the Other Zone). It was expected that all the moths  
158 exposed to the US during the test would lay more eggs on the Stimulus Zone than on the  
159 Other Zone, independently of previous conditioning (Davis & Stamps, 2004; Hopkins,  
160 1916). However, during the test with the CS, it was expected that Group Paired would  
161 lay more eggs in the area of the box where the CS had been placed (Stimulus Zone). To  
162 the contrary, in the presence of the CS, moths in the Unpaired or in the CS-Only groups  
163 were not expected to show such spatial preference for laying their eggs. That is, moths  
164 were not expected to prefer to lay eggs on the side that contains a stimulus, but only a  
165 stimulus that either promotes successful larvae (the mulberry leaves) or one associated  
166 with that stimulus as could occur in Group Paired.

167 Method

168 *Subjects*

169 Subjects were 90 silkworm moths (*Bombyx mori*), 45 males and 45 females. The  
170 female moths were randomly assigned to three groups: Paired (N=16), Unpaired (N=16)

171 and CS-Only (N=13). The larvae were purchased from a vendor located through the  
172 internet ( <https://www.milanuncios.com/anuncios/gusanos-de-seda.htm>).

### 173 *Apparatus*

174 For this experiment four cardboard boxes measuring 33 x 18 x 11 cm (*H x W x D*)  
175 were used to house the moths, one of them for male moths and the others for each group  
176 of female moths. Two mulberry leaves (*Morus alba*) were used as the US. Three drops  
177 (approximately 0.15 ml) of fresh cologne (VéGé) were used as the CS.

178 Transparent plastic boxes measuring 13 x 13 x 8 cm were used for training. The  
179 CS was contained in a 3 cm diameter circular plastic container. For testing three  
180 transparent plastic boxes measuring 32 x 16 x 12 cm were used.

### 181 *Procedure*

182 In their larval stage the subjects remained in cardboard boxes, where they were  
183 fed with mulberry leaves three times a day until they began metamorphosis. After  
184 hatching, males and females were separated into different boxes. Males remained  
185 together in a box until the test. Females were randomly assigned to one of the three  
186 groups and separated into different boxes by group.

187 The experiment consisted of two phases: conditioning and test.

188 *Conditioning.* Female moths received 6 sessions of conditioning with 5 trials each  
189 session. After each conditioning trial there was a 3 minute inter-trial interval (ITI).  
190 Moths were trained individually or in groups of 2-5 animals, depending on how many  
191 moths were ready. The moths conditioned at the same time belonged to the same group.  
192 The first session was the first afternoon after hatching. On the next two days moths  
193 received a session in the morning (around 9:00 h) and another one in the afternoon



194 (around 21:00 h). Last session was the morning of the fourth day. The central column of  
195 Table 1 shows the treatment received by each group during conditioning.

196 The moths in Paired Group experienced the CS and the US together for 5 minutes  
197 on each trial (6 sessions x 5 trials x 5 minutes = 150 minutes of experience with each  
198 stimulus). On a trial the stimuli were presented in the middle of the box with the CS  
199 presented in the small dish and the US presented by placing two mulberry leaves next to  
200 the CS. The CS and US were not changed between trials or between moths in a session.  
201 The moths were placed in the center of the box and allowed to freely behave for 5  
202 minutes with the box top covered.

203 For Unpaired Group, moths in each trial were exposed to CS or US (half of the  
204 subjects started with the CS and the other half started with the US). The stimulus  
205 sequence used over the six sessions in this group was CS-US-US-CS-CS-US or US-CS-  
206 CS-US-US-CS, so that they received 3 sessions with CS and 3 sessions with US. Each  
207 subject in this group was randomly assigned to one of the two sequences. Trials in this  
208 group lasted 10 minutes so that the exposure time of these subjects to stimuli was the  
209 same as in the Paired Group (3 sessions by stimulus x 5 trials x 10 minutes = 150  
210 minutes of experience with each stimulus). Finally, the CS-Only Group received the  
211 same treatment as Paired Group, with the exception that only the CS was presented in  
212 the conditioning box. The stimuli during conditioning were placed in a central position  
213 on the ground of the conditioning box in the same way as in the Paired group. Between  
214 trials the moths were removed and spent a 3-min ITI in boxes identical to the  
215 conditioning boxes, but these were completely empty.

216 *Test.* This phase began the afternoon of the fourth day after hatching and lasted 24  
217 hours. The test started by placing a male and a female moth in the center of the test box.  
218 After this time, the moths were removed, and eggs were counted, differentiating those

219 laid on the Stimulus Zone of those laid on the Other Zone (see Figure 1). In this phase  
220 half of the moths of each group were exposed only to the US and the other half only to  
221 the CS, so that in half of the cases Stimulus Zone represents the area where the US has  
222 been presented, whereas in the other half the CS was the stimulus placed on that area.  
223 During the test the US or CS were placed in a central position in the Stimulus Zone.

#### 224 *Statistical Analysis*

225 The dependent variable was the percentage of eggs laid in each zone in the test  
226 box. This percentage was calculated as indicated by the formula below.

$$227 \frac{\text{number of eggs in Stimulus Zone}}{\text{number of eggs in Stimulus Zone} + \text{number of eggs in Other Zone}} \times 100$$

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229 The mean percentage of eggs laid in each zone was compared using analyses of  
230 variance (ANOVA). The rejection criterion was set at  $p < .05$ , and effect sizes were  
231 reported using partial eta-squared ( $\eta_p^2$ ). Moreover, 90% confidence intervals for the  
232 effect sizes were calculated and reported for each analysis using the software provided  
233 by Nelson (2016).

#### 234 Results and Discussion

235 Figure 2 shows the mean percentage of eggs laid by each group during the test on  
236 the Stimulus and Other Zones broken down by the test stimulus (CS or US). Black bars  
237 show the mean percentage of eggs laid on the Stimulus Zone and white bars indicate the  
238 mean percentage of eggs laid on the Other Zone.

239 As it can be seen in the figure, all groups laid a higher percentage of eggs in the  
240 Stimulus Zone when the US was presented. However, when the CS was presented,  
241 differences between groups appeared. To analyze the patterns shown in the graphs a 3  
242 Group (Paired vs. Unpaired vs. CS-Only) x 2 Stimulus (CS vs. US) x 2 Zone (Stimulus

243 Zone vs. Other Zone) ANOVA was conducted. This analysis found a significant main  
244 effect of Zone,  $F_{1, 39} = 66.14$ ,  $P < .001$ ,  $\eta_p^2 = .62$  [CI: .46- .72], as well as the two-way  
245 interactions Group x Zone  $F_{2, 39} = 7.36$ ,  $P < .01$ ,  $\eta_p^2 = .27$  [CI: .07- .42], and Stimulus x  
246 Zone,  $F_{1, 39} = 21.96$ ,  $P < .001$ ,  $\eta_p^2 = .36$  [CI: .16- .51] and, more interestingly, the Group  
247 x Stimulus x Zone three-way interaction,  $F_{2, 39} = 6.65$ ,  $P < .01$ ,  $\eta_p^2 = .25$  [CI: .06- .40].

248 To analyze the 3-way interaction we conducted Stimulus x Zone ANOVAs within  
249 each group. In the Paired Group, there was an effect of Zone  $F_{1, 14} = 294.66$ ,  $P < .001$ ,  
250  $\eta_p^2 = .95$  [CI: .89- .97] and no interaction,  $F < 1$ . There were significant, and large,  
251 effects of zone both the US stimulus  $F_{1, 7} = 110.25$ ,  $P < .001$ ,  $\eta_p^2 = .94$  [CI: .77- .96],  
252 and, of most importance, the CS stimulus  $F_{1, 7} = 215.54$ ,  $P < .001$ ,  $\eta_p^2 = .97$  [CI: .87-  
253 .98]

254 The same analysis focused on the Unpaired Group found a Stimulus x Zone  
255 interaction,  $F_{1, 14} = 16.45$ ,  $P < .001$ ,  $\eta_p^2 = .54$  [CI: .19- .70], although the main effect of  
256 Zone approached significance,  $F_{1, 14} = 4.31$ ,  $P = .057$ ,  $\eta_p^2 = .23$  [CI: .00- .48]. The effect  
257 of Zone was significant when the US was present,  $F_{1, 7} = 94.99$ ,  $P < .001$ ,  $\eta_p^2 = .93$  [CI:  
258 .74- .96], but not when CS was present,  $F_{1, 7} = 1.08$ ,  $P = .332$ ,  $\eta_p^2 = .13$  [CI: .00- .45],  
259 indicating the moths in the Unpaired Group laid a similar percentage of eggs on both  
260 zones when CS was present during the test.

261 A similar pattern was observed in the CS-Only Group, where there was a main  
262 effect of Zone,  $F_{1, 11} = 10.23$ ,  $P < .01$ ,  $\eta_p^2 = .48$  [CI: .09- .67], and a Stimulus x Zone  
263 interaction,  $F_{1, 11} = 7.59$ ,  $P = .019$ ,  $\eta_p^2 = .40$  [CI: .05- .62]. The simple effect of Zone was  
264 significant when the US was present,  $F_{1, 5} = 98.36$ ,  $P < .001$ ,  $\eta_p^2 = .95$  [CI: .74- .97], but  
265 not with the CS,  $F < 1$ . As in the Unpaired Group, moths laid a higher percentage of eggs

266 on the Stimulus Zone than the Other Zone when the US was present, with no  
267 differences between the zones when only the CS was present.

268 As statistical analyses have confirmed, regardless of whether the moths had been  
269 exposed to the uncorrelated CS and US, only to the CS, or to the paired CS-US  
270 presentations, in the presence of the US all the moths preferred to lay the eggs near that  
271 stimulus (mulberry leaves). This result is consistent with the Hopkins principle  
272 (Hopkins, 1916) and the natal habitat preference induction hypothesis (Davis & Stamps,  
273 2004). It is possible that the moths had laid the eggs near the mulberry (the host plant)  
274 because it was the plant they ate when they were larvae or simply because it is the only  
275 plant present in the box. An additional experiment would be needed to determine the  
276 merits of that possibility allowing the moths the possibility of choosing between the  
277 host plant and a non-host plant during the oviposition test.

278 Nevertheless, when the CS was present in the test, the results were different  
279 depending on the group. In the Paired Group the percentage of eggs was higher in  
280 Stimulus Zone than in Other Zone in presence of the CS. That is, moths that  
281 experienced the odor paired with the mulberry leaves preferred to lay their eggs close to  
282 the odor when it was present. The oviposition choices were conditioned. When the CS  
283 was present, the difference between the number of eggs on the Stimulus and Other  
284 zones was not reliable in the Unpaired Group and CS-Only Groups. That the difference  
285 appeared only in the group for which the leaves and odor were paired supports that the  
286 result found in Paired Group is due to conditioning.

287 In absence of mulberry leaves, moths prefer to lay their eggs near something, the  
288 odor in this case, that had been previously associated with that plant. As we said above,  
289 the silkworm does not eat during the adult phase, so the influence of the moth's food  
290 preferences on the oviposition behavior can be precluded. To our knowledge, this is the

291 first clear evidence of learning in an insect species that is economically important as the  
292 producer of silk. Moreover, our results support the statement that post-hatching  
293 experience can affect oviposition choice (e.g., Olsson et al., 2006), and that one  
294 important source of this experience comes from a basic learning process such as  
295 Pavlovian conditioning. Thus, silkworms not only are able to use associative learning,  
296 but this learning can have an important biological function in this insect.

297         The behavior studied here is very relevant for a lepidopteran from an ecological  
298 point of view. Though the adaptive value of conditioning has been demonstrated in  
299 many species (e.g., Domjan et al., 1998; Guillette et al., 2009; Hollis, 1984), it should  
300 be noted that studies concerned with Pavlovian conditioning, contrary to the experiment  
301 presented here, often show little clear evidence regarding the adaptive value of the  
302 conditioned response acquired by the organism, human or otherwise. In the present  
303 research, Pavlovian conditioning contributes the foraging of offspring, thus the results  
304 reported here evidence that Pavlovian conditioning can provide a clear advantage to  
305 silkworms for the species' survival.

306         Finally, it should be noted that this experiment explored associative learning that  
307 has been acquired during the adult phase of the silkworm. Nevertheless, it would stand  
308 to reason that classical conditioning will be adaptive if the knowledge acquired by  
309 larvae can be used by moths. Such questions can be addressed with new research.

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## References

325 Adkins-Regan, E. & MacKillop, E. A. (2003). Japanese quail (*Coturnix japonica*)

326 inseminations are more likely to fertilize eggs in a context predicting mating

327 opportunities. *Proceedings of the Royal Society B*, 270, 1685–1689.

328 <http://dx.doi.org/10.1098/rspb.2003.242>

329 Alcock, J. (1993). *Animal Behaviour*. Sunderland, Massachusetts: Sinauer Associates.

330 Anderson, P., Hansson, B.S., Nilsson, U., Han, Q., Sjöholm, M., Skals, N. & Anton, S.

331 (2007). Increased behavioral and neuronal sensitivity to sex pheromone after brief

332 odor experience in a moth. *Chemical Senses*, 32, 483–491.

333 <http://dx.doi.org/10.1093/chemse/bjm017>

- 334 Anderson, P., Sadek, M.M., Larsson, M., Hansson, B.S. & Thöming, G. (2013). Larval  
335 host plant experience modulates both mate finding and oviposition choice in a  
336 moth. *Animal Behaviour*, 85, 1169–1175.  
337 <http://dx.doi.org/10.1016/j.anbehav.2013.03.00>
- 338 Barron, A.B. (2001). The life and death of Hopkins' host-selection principle. *Journal of*  
339 *Insect Behaviour*, 14, 725–737. <http://dx.doi.org/10.1023/A:1013033332535>
- 340 Blackiston, D.J., Casey, E.S. & Weiss, M.R. (2008). Retention of memory through  
341 metamorphosis: Can a moth remember what it learned as a caterpillar? *PLOS*  
342 *ONE*, 3 (3), e1736. <http://dx.doi.org/10.1371/journal.pone.0001736>
- 343 Coria-Avila, G.A. (2012). The role of conditioning on heterosexual and homosexual  
344 partner preferences in rats. *Socioaffective Neuroscience & Psychology*, 2, 17340.  
345 <http://dx.doi.org/10.3402/snp.v2i0.17340>
- 346 Cunningham, J. P., Jallow, M. F. A., Wright, D. J. & Zalucki, M. P. 1998. Learning in  
347 host selection in *Helicoverpa armigera* (Hubner) (Lepidoptera: Noctuidae).  
348 *Animal Behaviour*, 55, 227-234. <http://dx.doi.org/10.1006/anbe.1997.0600>
- 349 Cunningham, J.P., Moore, C.J., Zalucki, M.P. & West, S.A. (2004). Learning, odor  
350 preference and flower foraging in moths. *Journal of Experimental Biology*, 207,  
351 87–94. <http://dx.doi.org/10.1242/jeb.00733>
- 352 Daly, K.C. & Smith, B.H. (2000). *Associative olfactory learning in the moth Manduca*  
353 *sexta*. *Journal of Experimental Biology*, 203, 2025–2038.
- 354 Davis, J. M. & Stamps, J. (2004). The effect of natal experiences on habitat preferences.  
355 *Trends in Ecology & Evolution*, 19, 411-416.  
356 <http://dx.doi.org/10.1016/j.tree.2004.04.006>

357 Domjan, M., Blesbois, E., & Williams, J. (1998). The adaptive significance of sexual  
358 conditioning: Pavlovian control of sperm release. *Psychological Science*, 9, 411–  
359 415. <http://dx.doi.org/10.1111/1467-9280.00077>

360 Dukas, R. (1999). Ecological relevance of associative learning in fruit fly larvae.  
361 *Behavioural Ecology and Sociobiology*, 45, 195-200.  
362 <http://dx.doi.org/10.1007/s002650050553>

363 Dukas, R. (2006). Learning in the context of sexual behavior in insects. *Animal*  
364 *Behaviour*, 56, 125–141. <http://dx.doi.org/10.1163/157075606777304258>

365 Dukas, R. & Bernays, E.A. (2000). Learning improves growth rate in grasshoppers.  
366 *Proceedings of the National Academy of Sciences*, 97, 2637–2640.  
367 <http://dx.doi.org/10.1073/pnas.050461497>

368 Fan, R.J., Anderson, P. & Hansson, B.S. (1997). Behavioural analysis of olfactory  
369 conditioning in the moth *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae).  
370 *Journal of Experimental Biology*, 200, 2969–2976.

371 Gandolfi, M., Mattiacci, L. & Dorn, S. (2003). Preimaginal learning determines adult  
372 response to chemical stimuli in a parasitic wasp. *Proceedings of the Royal Society*  
373 *B*, 270, 2623–29. <http://dx.doi.org/10.1098/rspb.2003.2541>

374 Guillette, L.M., Hollis, K.L. & Markarian, A. (2009). Learning in a sedentary insect  
375 predator: antlions (Neuroptera: myrmeleontidae) anticipate a long wait.  
376 *Behavioural Processes*, 80, 224–232.  
377 <http://dx.doi.org/10.1016/j.beproc.2008.12.015>

378 Hollis, K. L. (1984). The biological function of Pavlovian conditioning: the best defense  
379 is a good offense. *Journal of Experimental Psychology: Animal Behaviour*  
380 *Processes*, 10, 413-425. <http://dx.doi.org/10.1037/0097-7403.10.4.413>



381 Hollis, K.L., Dumas, M.J., Singh, P. & Fackelman, P. (1995). Pavlovian conditioning of  
382 aggressive behavior in blue gourami (*Trichogaster trichopterus*): winners become  
383 winners and losers stay losers. *Journal of Comparative Psychology*, 109, 123–  
384 133. <http://dx.doi.org/10.1037/0735-7036.109.2.123>

385 Hollis, K.L., Pharr, V.L., Dumas, M.J., Britton, G.B. & Field, J. (1997). Classical  
386 conditioning provides paternity advantage for territorial male blue gouramis  
387 (*Trichogaster trichopterus*). *Journal of Comparative Psychology*, 111, 219–225.  
388 <http://dx.doi.org/10.1037/0735-7036.111.3.219>

389 Hopkins, A.D. (1916). Economic investigations of the scolytid bark and timber beetles  
390 of North America. *U.S. Department of Agriculture Program of Work for 1917*.

391 Janz, N., Söderlind, L. & Nylin, S. (2008). No effect of larval experience on adult host  
392 preferences in *Polygonia c-album* (Lepidoptera: Nymphalidae): on the persistence  
393 of Hopkins' host selection principle. *Ecological Entomology*, 34, 50–57.  
394 <http://dx.doi.org/10.1111/j.1365-2311.2008.01041.x>

395 Jones, P.L. & Agrawal, A.A. (2017). Learning in Insect Pollinators and Herbivores.  
396 *Annual Review of Entomology*, 62, 53-71. [http://dx.doi.org/10.1146/annurev-ento-](http://dx.doi.org/10.1146/annurev-ento-031616-034903)  
397 [031616-034903](http://dx.doi.org/10.1146/annurev-ento-031616-034903)

398 Jørgensen, K., Strandén, M., Sandoz, J.C., Menzel, R. & Mustaparta, H. (2007). Effects  
399 of two bitter substances on olfactory conditioning in the moth *Heliothis virescens*.  
400 *Journal of Experimental Biology*, 210, 2563–2573.  
401 <http://dx.doi.org/10.1242/jeb.004283>

402 Nelson, J. B. (2016). A robust function to return the cumulative density of non-central F  
403 distributions in Microsoft Office Excel. *Psicologica: International Journal of*  
404 *Methodology and Experimental Psychology*, 37(1), 61-83. Retrieved from  
405 <https://www.uv.es/psicologica/articulos1.16/4NELSON.pdf>

- 406 Olsson, P.O.C., Anderbrant, O. & Löfstedt, C. (2006). Experience influences  
407 oviposition behaviour in two pyralid moths, *Ephestia cautella* and *Plodia*  
408 *interpunctella*. *Animal Behaviour*, 72, 545–551.  
409 <http://dx.doi.org/10.1016/j.anbehav.2005.10.023>
- 410 Papaj, D.R. (1986). Conditioning of leaf-shape discrimination by chemical cues in the  
411 butterfly, *Battus philenor*. *Animal Behaviour*, 34, 1281–1288.  
412 [http://dx.doi.org/10.1016/S0003-3472\(86\)80199-3](http://dx.doi.org/10.1016/S0003-3472(86)80199-3)
- 413 Riffell, J.A., Alarcón, R., Abrell, L., Davidowitz, G., Bronstein, J.L. & Hildebrand, J.G.  
414 (2008). Behavioral consequences of innate preferences and olfactory learning in  
415 hawkmoth-flower interactions. *Proceedings of the National Academy of Sciences*  
416 *of the U.S.A.*, 105, 3404–3409. <http://dx.doi.org/10.1073/pnas.0709811105>
- 417 Singh, T. & Saratchandra, B. (2004). *Principles and Techniques of Silkworm Seed*  
418 *Production*. New Delhi: DPH.
- 419 Thorpe, W.H. & Jones, F. G. W. (1937). Olfactory conditioning in a parasitic insect and  
420 its relation to the problem of host selection. *Proceedings of the Royal Society B*,  
421 124, 56-81. <http://dx.doi.org/10.1098/rspb.1937.0072>
- 422 Traynier, R. (1984). Associative learning in the ovipositional behaviour of the cabbage  
423 butterfly, *Pieris rapae*. *Physiological Entomology*, 9, 465-472.  
424 <http://dx.doi.org/10.1111/j.1365-3032.1984.tb00789.x>
- 425 Traynier, R. (1986). Visual learning in assays of sinigrin solution as an oviposition  
426 releaser for the cabbage butterfly, *Pieris rapae*. *Entomologia Experimentalis et*  
427 *Applicata*, 40, 25–33. <http://dx.doi.org/10.1007/BF00187019>
- 428 Tully, T., Cambiazo, V. & Kruse, L. (1994). Memory through metamorphosis in normal  
429 and mutant *Drosophila*. *Journal of Neuroscience*, 14, 68–74.  
430 <http://dx.doi.org/10.1523/JNEUROSCI.14-01-00068.1994>

431 Zamble, E., Hadad, G. M., Mitchell, J. B., & Cutmore, T. R. H. (1985). Pavlovian  
 432 conditioning of sexual arousal: First- and second-order effects. *Journal of*  
 433 *Experimental Psychology: Animal Behaviour Processes*, 11, 598–610.  
 434 <http://dx.doi.org/10.1037/0097-7403.11.4.598>

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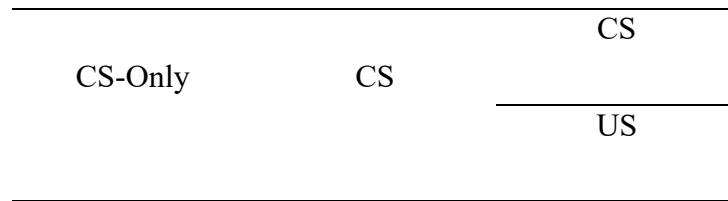
442 *Table 1. Experimental design*

443 CS: Conditioned stimulus (odor); US: Unconditioned stimulus (mulberry leaves).

444 Arrow indicates simultaneous presentation, whereas the slash indicates uncorrelated

445 presentations.

<b>Group</b>	<b>Training</b>	<b>Test</b>
Paired	CS → US	CS
		US
Unpaired	CS / US	CS
		US



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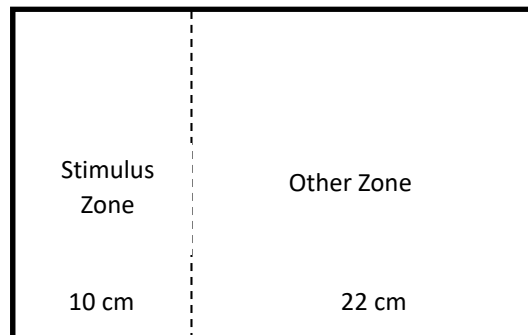
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459 *Figure 1.* Schematic of the Test box. “Stimulus Zone” is the space in which the stimulus

460 (CS or US) is presented during the test. “Other Zone” refers to the rest of the box.

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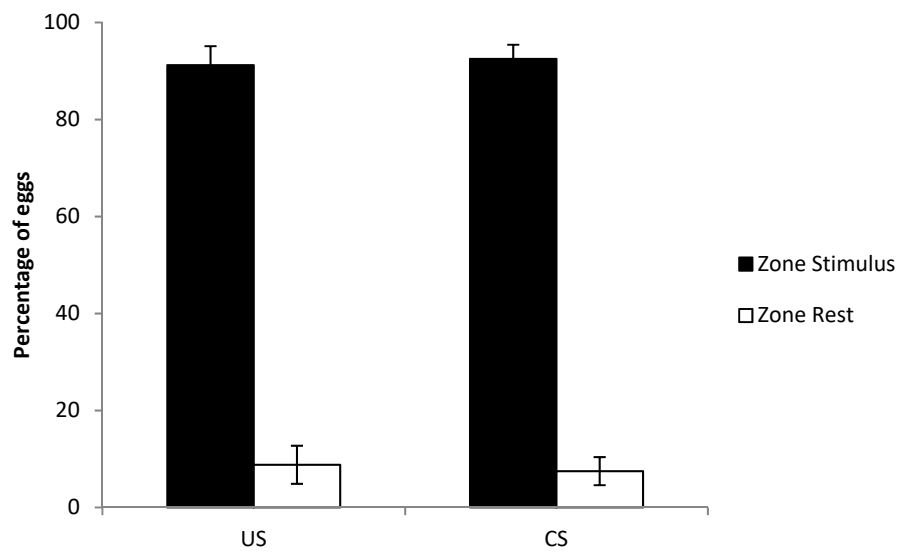
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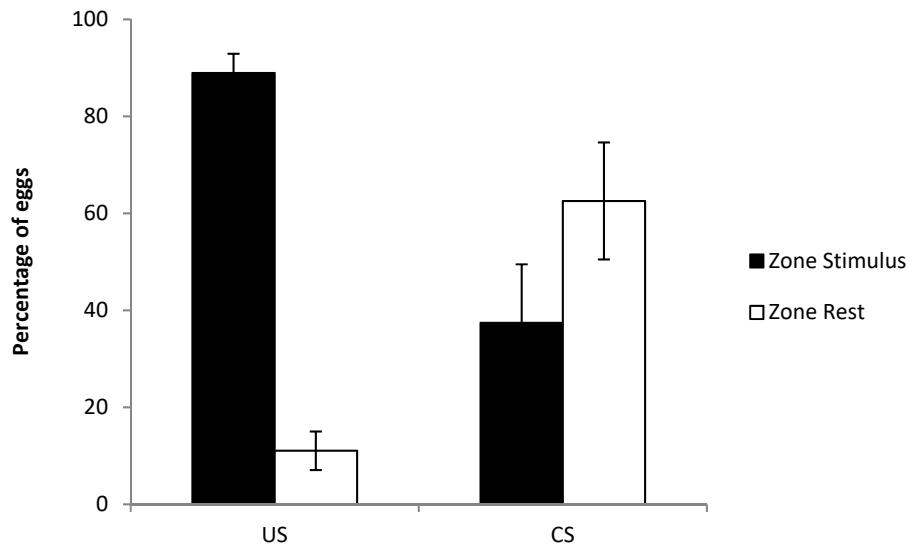
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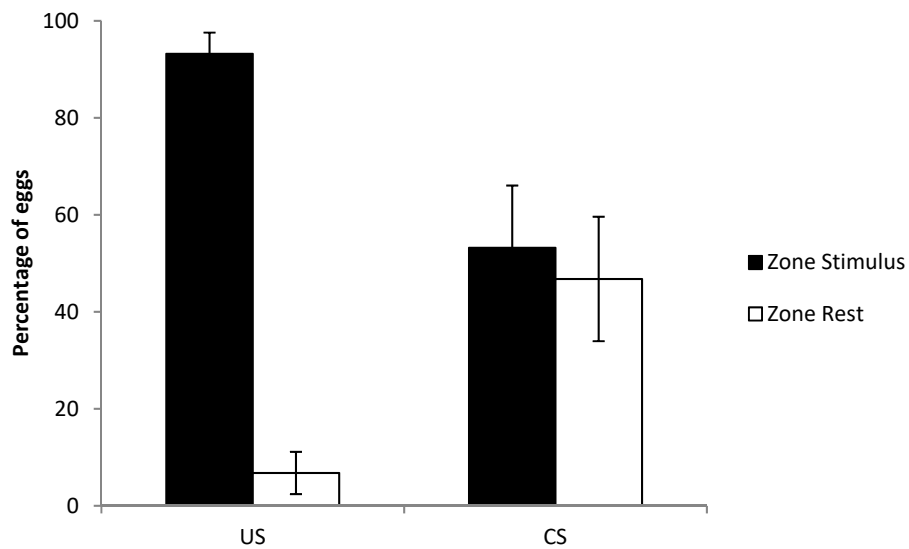


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487 C



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490 *Figure 2.* Percentage of eggs laid during the test in each box zone (Stimulus Zone or  
 491 Other Zone), depending on whether the stimulus present was the US or the CS.  
 492 Subjects of the Paired Group are shown in panel A, the Unpaired Group in panel  
 493 B, and the CS-Only Group in panel C). The left section of each graph shows the  
 494 results for the moths tested in the presence of the mulberry leaves (US), and the  
 495 right shows results for the moths tested in the presence of the odor (CS).

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